MYCOPLASMA

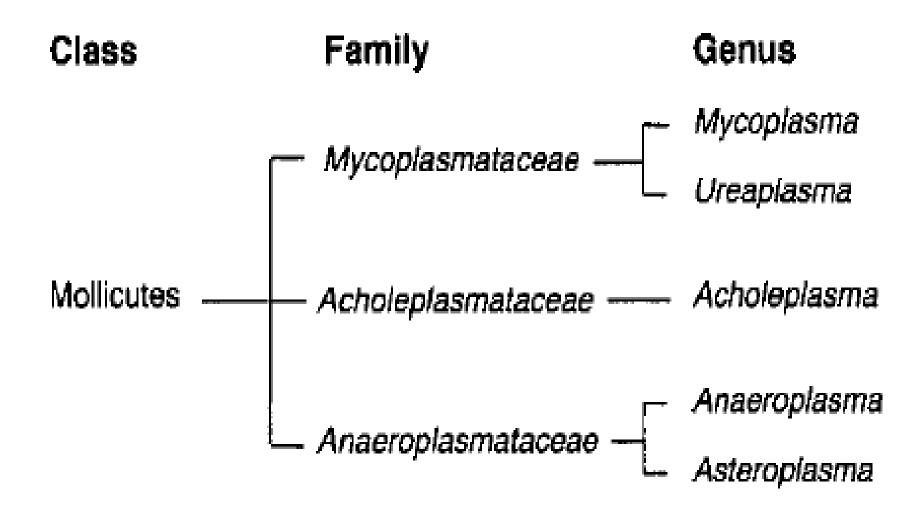
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General Characteristics

- Smallest prokaryotic cells capable of self-replication
- Cannot synthesize Peptidoglycan (resistant to penicillin)
- Do not possess rigid cell walls but have flexible, triple-layed outer membranes
- Lack rigid cell wall and thus definite shape (Pleomorphic organisms)
- Being pleomorphic, they can pass through bacterial filters of 0.22 μ m size.
- Require Sterol for growth (except-Acholeplasma)
- Earlier term *Pleuropneumonia- like organisms (PPLO)* was in use for mycoplasmal organisms.



Media for Mycoplasma

- Smallest prokaryotes that can be grown on artificial media.
- Fastidious in growth requirement.
- Requires **sterol** and animal protein for growth
- **Penicillin** inhibits growth of Gram positive bacteria
- Thallium acetate inhibits Gram negative bacteria and fungi.

Media for mycoplasma contains:

- Heart infusion broth
- 20% Horse serum
- 1% Peptone
- 1% Albumin
- Penicillin 1000 IU/ml
- -Thallium acetate
 - 1: 2000 to 1: 8000 conc.
- -Yeast extract

Culturing Mycoplasma

- Can be grown in liquid or solid media
- Facultative anaerobe require 5-10% CO₂
- Optimal temperature for growth 35-37°C
- Generally slow grower (generation interval up to 9hrs)
- For culturing ureaplasmas, urea is added to the medium and thallium acetate is removed, which is toxic for these organisms

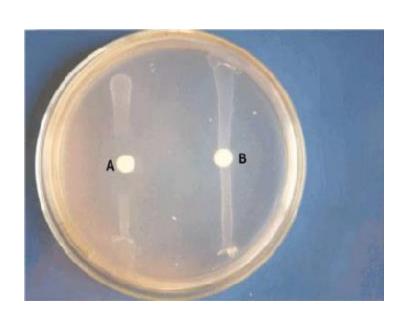
Mycoplasmal colonies

- Mycoplasma colonies are small and can only be observed under a light microscope at low magnification
- Dienes stain is used for staining mycoplasmal microcolonies
- Nipple Shaped colony or Umbonate micro-colonies – Fried egg appearance
- Because of tiny colonies of Ureaplasmathey were also called *T-mycoplasma*



Genus	Colony size
Mycoplama	0.1-0.6 mm
Ureaplasma	0.02-0.06 mm
Acholeplasma	Upto 1.5 mm

Requirement for Sterol and Urea hydrolysis



 Mycoplasmas requires Sterol except members of genus Acholeplasma.

 The requirement of sterol is ascertained performing "Digitonin" sensitivity.

- Digitonin impregnated discs placed on solid media inhibits growth of isolates that require sterol.
- Members of genus Ureaplasma produce *Urease* and thus capable of hydolysis "urea".

Isolate	Effect of digitonin	Requirement for cholesterol	Urease production	Colony size
Mycoplasma species	Growth inhibition	+	-	0.1 – 0.6 mm
Ureaplasma species	Growth inhibition	+	+	0.02 – 0.06 mm
Acholeplasma species	No growth inhibition	_	_	up to 1.5 mm

Mycoplasma and L form of bacteria

Mycoplasma and L type bacteria are similar:

- they both lack a cell wall and the cell is pleomorphic
- they can both pass through an antimicrobial filter

T	he main	differences	between t	he two are:

Mycoplasma	L –form of Bacteria
Mycoplasma are independent microbes	L-type bacteria are variants of normal bacterial cells that have a cell wall (most L-type cells will revert to their original
	form)
Mycoplasma growth requires cholesterol (10–20% serum in the medium)	L-type bacteria does not sterol
Mycoplasma do not fade easily after Dienes staining	L-type bacteria fade easily after Diane staining

Habitat

- Mycoplasmas are found on mucosal surfaces of the respiratory, intestinal and genital tracts
- Members of Anaeroplasma are strict anaerobes remain in rumen of cattle and sheep
- Members of Spiroplasma causes disease in plants.
- Factors such as extremes of age, stress and intercurrent infection may predispose disease condition.
- In addition, mycoplasmas may exacerbate disease initiated by other pathogens, particularly in the respiratory tract.
- Mycoplasmal infections cause respiratory diseases of major economic importance in farm animals especially in ruminants, pigs and poultry

Mycoplasma species	Hosts	Disease conditions
M. mycoides subsp. mycoides (small colony type)	Cattle	Contagious bovine pleuropneumonia
M. bovis	Cattle	Mastitis, pneumonia, arthritis
M. agalactiae	Sheep, goats	Contagious agalactia
M. capricolum subsp. capripneumoniae (F38)	Goats	Contagious caprine pleuropneumonia
M. capricolum subsp. capricolum	Sheep, goats	Septicaemia, mastitis, polyarthritis, pneu- monia
M. mycoides subsp. mycoides (large colony type)	Goats, sheep	Pleuropneumonia, mastitis, septicaemia, polyarthritis
M. mycoides subsp. capri	Goats	Septicaemia, pleuropneumonia, arthritis, mastitis

M. hyopneumoniae	Pigs	Enzootic pneumonia
M. hyorhinis	Pigs (3-10 weeks of age)	Polyserositis
M. hyosynoviae	Pigs (10-30 weeks of age)	Polyarthritis
M. gallisepticum	Chickens Turkeys	Chronic respiratory disease Infectious sinusitis
M. synoviae	Poultry	Infectious synovitis
M. meleagridis	Turkeys	Airsacculitis, bone deformities, reduced hatchability and growth rate

Contagious Bovine Pleuro-pneumonia (CBPP)

A severe contagious disease of cattle caused by M.
 mycoides subspecies mycoides (small colony type)

Transmission is by aerosols- requires close contact

In severe outbreaks the mortality rate may be high

 Death can occur 1 to 3 weeks after the onset of clinical signs.

CBPP- Symptoms

- Acute form sudden onset of high fever, anorexia, depression, drop in milk yield, accelerated respiration and coughing.
- Animals adopt a characteristic stance with the head and neck extended and elbows abducted.
- Expiratory grunting and mucopurulent nasal discharge may be present.
- Arthritis, synovitis and endocarditis may be present in affected calves.





CBPP- PM findings

- Pneumonic lungs have a marbled appearance.
- Grey and red consolidated lobules
- Interlobular septa are distended and oedematous.
- Serofibrinous exudate in the pleural cavity.
- In chronic cases, *fibrous encapsulation of necrotic foci* is commonly found.
- These necrotic foci contain viable mycoplasmas





CBPP- Diagnosis and Control

Diagnosis

- Clinical signs and characteristic PM findings helps in presumptive diagnosis.
- PCR, FAT can be used on pleural fluid to confirm the presence of the pathogen.
- Broncho-alveolar lavage, pleural fluid, lung tissue or the bronchopulmonary lymph nodes

Serological tests:

- Rapid field serum agglutination test
- Passive haemagglutination screening test
- -Complement fixation test –
- Dot-blot technique for confirmation

Treatment and control

Treatment is generally unsatisfactory

Pneumonia, Mastitis, Polyarthritis

- Mycoplasma bovis causes severe pneumonia in calves
- It can exacerbate respiratory disease caused by Pasteurella and Mannheimia species
- · Most important mycoplasmal species causing mastitis.
- Mycoplasmal mastitis is common in large daily herds.
- Milking machines, milkers' hands and cloths which are then important sources of infection Causes severe mastitis, systemic involvement is uncommon.
- subclinical carriers are important sources of infection
- Dramatic loss of milk production and the serous or purulent mastitic exudates
- Infection often results in agalactia

Mycoplasmal infections in Goats

Contagious caprine pleuropneumonia	 Caused by M. capricolum subspecies capripneumoniae (Mycoplasma strain F38) Similar in many respects to CBPP Highly contagious disease; transmission through aerosols. The disease is characterized by pneumonia, fibrinous pleurisy, profuse pleural exudate Marbled appearance on the cut surface of affected lungs is seen. Inactivated vaccines give satisfactory protection. Pleuropneumonia in goats can occasionally be caused by M. mycoides subspecies capri M. mycoides subspecies mycoides (large colony type) M. capricolum subspecies capricolum
Contagious agalactia of sheep and goats Mastitis, Pneumonia, Arthritis	 Caused by M. agalactiae Characterized by mastitis, arthritis and conjunctivitis. Observed soon after parturition Pregnant animals may abort Can be fatal in young animals due to pneumonic complications. The organism is shed in milk Organism may remain localized in the supra-mammary lymph nodes M. mycoides subspecies capri M. mycoides subspecies mycoides (large colony type) M. capricolum subspecies capricolum

Mycoplasmal infections in Pigs

Enzootic pneumonia	 Caused by M. hyopneumoniae, Affects intensively reared pigs. Extremes of temperature, sudden change in temperature, poor ventilation, overcrowding Pigs of all ages are susceptible Characterized by coughing, poor growth rates and, in some cases, respiratory distress. At post mortem, pulmonary consolidation is confined to the apical and cardiac lobes with clear demarcation from normal lung tissue.
Polyserosititis	 Caused by Mycoplasma hyorhinis Chronic progressive polyserositis in pigs up to 10 weeks of age. It is characterized by fever, laboured breathing, lameness and swollen joints.
Polyarthritis	 Caused by M. hyosynoviae Affects pigs from 10 to 30 weeks of age. This self-limiting arthritis and synovitis produces transient lameness.

Poultry infections

 Mycoplasma gallisepticum causes chronic respiratory disease in chickens and infectious sinusitis in turkeys.

Transmission:

- Through infection of the embryo in the egg
- By aerosols
- <u>In chickens:</u> affects upper respiratory tract.
- <u>In turkeys:</u> Causes swelling of the paranasal sinuses

Diagnosis:

- Isolation and identification of the pathogen
- Flock testing using the serum plate agglutination test
- Haemagglutination inhibition and ELISA tests
- Specific-pathogen-free (SPF) flocks
- Eggs used for hatching should be dipped in a tylosin

Poultry infections

Mycoplasma gallisepticum	Chronic respiratory disease in chickens Infectious sinusitis in turkeys
Mycoplasma meleagridis	Causes airsacculitis in young poults and joint and bone deformities in growers Transmission is mainly through eggs
Mycoplasma synoviae	Infectious synovitis - In chickens and turkeys -Transmitted mainly by aerosols. - Arthritis and respiratory signs are the main clinical features

Diagnosis of Mycoplasmal infections

- Suitable samples include mucosal scrapings, tracheal exudates, aspirates, pneumonic tissue, mastitic milk and fluids from joints or body cavities.
- The presence of Mycoplasma in samples can be demonstrated by:
 - -Fluorescent antibody techniques
 - -Peroxidase-antiperoxidase procedures on paraffin embedded tissues
 - -Polymerase chain reaction techniques

Diagnosis of Mycoplasmal infections

- Isolation can be attempted in mycoplasma medium under 10% CO₂ at 37° C for upto 14 days.
- Identification criteria for isolates:
 - 'Fried-egg' microcolonies
 - Microcolony size
 - -Cholesterol requirement for growth (digitonin sensitivity test)
 - -Biochemical profile including urease production
 - -Fluorescent antibody technique on microcolonies
 - -Growth inhibition test with specific antisera

Serological tests

- Complement fixation tests

-Tests based on ELISA

-Rapid plate agglutination tests

-Haemagglutination-inhibition tests

THANKS